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Screening of *in vitro* cytotoxic activity of *Ephedra alata* used traditionally to treat cancer in Libya

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Abstract

Ephedra is one of the Libyan medicinal herbs that are traditionally involved in folk medicine to treat several diseases. *Ephedra* is known to have antioxidant and antibacterial properties. The goal of this study is to evaluate the cytotoxic activity of methanolic extract of *Ephedra alata* plant growing wild in the east of Libya against two human cancer cell lines HEPG2 human liver cancer cell line and PC3 human prostate cancer cell line. The results show that the plant methanolic extract has a significant cytotoxic effect on both cell lines compared with doxorubicin as a positive control.

Keywords: *Ephedra alata*, human liver cancer cell line, human prostate cancer cell line and methanolic extract

1. Introduction

Research projects regarding cancer have evolved in the past decades since cancer is now the most common cause of death worldwide, making it of great concern among the world.

American Cancer Society has released global cancer statistics that by 2050 there is a prediction of 27 million new cancer cases and 17.5 million cancer deaths [1]. Various approaches have been developed to minimize the effect of cancer on the human body. In modern cancer treatment approaches, chemotherapy has become the leading acknowledged choice of therapy, along with other clinically available anti-cancer drugs, of synthetic or natural product origin, that are currently being utilized for the treatment of some types of cancer [2, 3].

In the Arab society, including the Libyan society, traditional herbal medicine has always been a way to cure a common minor illness such as a cough, constipation, diarrhea, stomach cramps, rheumatism, etc. Therefore, it has recently become a topic of interest in experimental fields and research. Quite a lot of studies have been conducted on herbs for their anti-cancer properties. For example, Hartwell has collected several data, approximately 3,000 plants, possessing anti-cancer properties, and later on used as potent cytotoxic drugs [4-6]. The secondary metabolites and their semi-synthetic derivatives obtained from the plants play an important role in anti-cancer drug therapy [7, 8]. *Ephedra* is one of these medicinal plants, belonging to the *Ephedraceae* family. Different species of *Ephedra* are distributed around the world; among these are *Ephedra Lristanica*, *Ephedra strobiliacea*, *Ephedra sarcocarpa*, *Ephedra procera*, *Ephedra pachyclada* and *Ephedra alata*, [9]. The herb grows abundantly in Libya, and used in traditional remedies for the treatment of chills, bronchial asthma, allergies, colds, edema, fever, coughs, flu, and headaches. This plant also shows anti-cancer properties [10-12]. Sympathomimetic alkaloids derived from phenylalanine such as ephedrine, pseudoephedrine, and other related compounds are the main components of *Ephedra alata* [13]. Several researches have proved that the aerial parts of this plant contain (-) ephedrine as the main isomer (between 30% and 90%) [14]. Additionally, *Ephedra* is considered a source of phenolic compounds such as trans-cinnamic acid, catechin, syringin, epicatechin, symplocoside, kaempferol 3-O-rhamnoside 7-O-glucoside and isovitexin 2-O-rhamnoside, which all contribute significantly to the antioxidant activity of the plant and play important roles in disease resistance [15-17]. Phenolic compounds are also believed to display an essential role as health-protecting factors. Scientific evidence indicates that consuming diets, which are rich in antioxidant compounds, will lower the risk of chronic diseases, including heart malfunction and cancer [18].

A universal property of tumor cells is the uncontrolled proliferation. Investigation of the cellular growth control mechanism has contributed to the understanding of carcinogenesis and the identification of compounds with specific anti-tumoral activity [19, 20].

Therefore, this study was designed to further insight and prove medical traditional uses of *Ephedra Alata* for anti-cancer remedies.

2. Material and methods

2.1 Plant material

The aerial parts of the *Ephedra Alata* were collected from Botraba area (about 100 km east of Benghazi, Libya) during August. Samples of the plant were sent to the Department of Pharmacognosy (Faculty of Pharmacy, University of Benghazi) for identification. The aerial parts were left to dry in open air. The dried plant materials were milled into a fine powder to be used for extraction.

2.2 Extraction preparation

The dried plant powder was extracted by cold maceration in 70% methanol. The solvent was then evaporated at a temperature of 40°C under reduced pressure. The residuals were stored for further biological or chemical examination.

2.3 Cytotoxic effect assessment

Cell viability was evaluated according to (Mosmann, 1983) through the mitochondrial-dependent reduction of the color of MTT (3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide) from yellow to purple [21].

The entire procedure was carried out in a sterile place using a Laminar flow cabinet biosafety class II level (Baker, SG403INT, and Sanford, ME, USA). The cells were suspended in DMEM medium, 1% antibiotic-antimycotic mixture (10,000µg/ml Streptomycin Sulfate, 10,000U/ml Potassium Penicillin, and 25µg/ml Amphotericin B) and 1% L-glutamine at a temperature of 37°C under 5% carbon dioxide.

The cells were batch cultured for ten days, then seeded in fresh complete growth medium in 96-well microtiter plastic plates at a concentration of 10×10^3 cells/well at 37°C for one day under 5% CO₂ using a water jacketed Carbon dioxide incubator (Sheldon, TC2323, Cornelius, OR, USA). The media was aspirated, fresh medium free of serum was added, the cells were incubated with different concentrations of the sample (100, 50, 25, 12.5, 6.25, 3.125, and 1.56µg/ml). Cells were incubated alone as a negative control. After two days of incubation, the medium was aspirated, 40µl MTT salt (2.5µg/ml) was added to each well and incubated for additional four hours at temperature of 37°C under 5% carbon dioxide. 200µL of 10% Sodium dodecyl sulphate in deionized water was added to each well and incubated at 37°C overnight to eliminate the reaction and dissolve of the formed crystals. A positive control composed of 100µg/ml doxorubicin was used as a standard cytotoxic natural agent that exerts 100% lethality under the same experimental conditions. The absorbance was measured at 595nm and a reference wavelength of 620nm by a micro plate multi-well reader (Bio-Rad Laboratories Inc., model 3350, Hercules, California, USA) [22-24].

SPSS 11 program was used to run independent t-test to evaluate the statistical significance between the negative control (cells with vehicle) and samples. DMSO (dimethyl sulfoxide) was used for the dissolution of the plant extracts and its final concentration on the cells was less than 0.2%. The percentage of change in viability was calculated using the following formula:

$$\left(\frac{\text{Reading of extract}}{\text{Reading of negative control}} - 1 \right) \times 100$$

3. Results and Discussion

The following table shows the cytotoxic activity of the plant extract against HepG2 human liver cancer cell line and PC3 human prostate cancer cell line.

Table 1: Cytotoxic Activity of the methanolic extract of the *Ephedra alata* (IC₅₀ µg/ml)*.

Drug name	HepG2 IC ₅₀	PC3 IC ₅₀
<i>Ephedra alata</i>	32.9	30.4
Doxorubicin	21.6	23.8

*Significance different from standard (Doxorubicin) $P < 0.05$ n=3

Cytotoxic effects of the extract of *Ephedra alata* was expressed as IC₅₀ values. According to the table, there was significant cytotoxicity on HepG2 and PC3 with IC₅₀; 32.9 and 30.4, respectively.

It is of interest that the extract of the plant showed cytotoxicity against the cancer cell lines, and if there would be corresponding results *in vivo*, then traditional healers would have some sort of scientific support for treating cancer patients.

Pathological examination of *E. pachyclada*, a different species from the same plant, showed that hepatic injuries, such as inflammation, necrosis and hepatitis, are partially healed with the plant extracts due to its hepato-protective mechanism to inhibit oxidative stress and inflammation [25].

The extracts of *Ephedra* significantly alleviate hepatocyte apoptosis and inflammatory factor infiltration by lowering of serum alanine aminotransferase and total bilirubin (T. Bil) activity, it also reduces the levels of TNF-α and the activities of caspase-3, 8, and 9 [26, 27].

A study has reported that there are anti-proliferative and analgesic effects in herbacetin, the aglycon of herbacetin 7-O-neohesperidoside isolated from *Ephedra* plants [28].

4. Conclusion

Several plant species rich in flavonoids are reported to have disease preventive and therapeutic properties. This observation is of particular significance since flavonoids are considered ingredients found in many fruits and vegetables, explaining the association of fruit and vegetable consumption with reduced cancer risk that have been reported in previous studies [29-31]. Cytotoxic activity recorded in this study collate with these findings since the phytochemical evaluation indicated the presence of flavonoids in the plant [16]. From the result, it is clear that the extract has potent *in vitro* cytotoxic activity against both cell lines. Further studies are also in a process to evaluate the most potent fraction of the active plant.

Furthermore, other parts of the *Ephedra* plant are also important to be further examined for their antimicrobial, antioxidant and cytotoxic effect.

5. References

1. American Cancer Society, Global Cancer Facts and Figures. American Cancer Society, Atlanta, 2007.
2. Chabner BA, Amrein PC, Druker BJ, Michaelson MD, Mitsiades CS, Gross PE *et al.* The Pharmacological Basis of Therapeutics. 11th ed. McGraw-Hill, New York, 1996, 1315-1403.
3. DeVita VT, Hellman S, Rosenberg SA. (Eds.), Cancer: Principles and Practice of Oncology. 8th ed. Lippincott-Williams & Wilkins Philadelphia, 2008.
4. Graham JG, Quinn ML, Fabricant DS, Farnsworth NR. Plants used against cancer-an extension of the work of Jonathan Hartwell. J Ethnopharmacol. 2000; 73(3):347-377.
5. Cragg GM, Newman DJ. Natural product scaffolds as leads to drugs. Future Med Chem. 2009; 1(8):1415-1427.
6. Shoeb M. Anti-cancer agents from medicinal plants.

- Bangladesh. J of Pharmacol. 2006; 1(2):35-41.
7. Pan L, Chai H, Kinghorn AD. Th continuing search for antitumor agents from higher plants. *Phytochem Lett.* 2010; 3(1):1-8.
 8. Indap MA, Radhika S, Motiwale L, Rao KVK. Quercetin: antitumor activity and pharmacological manifestations for increased therapeutic gains. *Indian J Pharm Sci.* 2006; 68:465-469.
 9. Rustaiyan A, Javidnia K, Farjam MH, Mohammadi MK, Mohammadi N. Total phenols, antioxidant potential and antimicrobial activity of the methanolic extracts of *Ephedra laristanica*. *Journal of Medicinal Plants Research.* 2011; 5(24):5713-5717.
 10. Konar RN, Singh MN. Production of plantlets from female gametophytes of *Ephedra foliata* Boiss. *Z. Pflanzenphysiol.* 1979; 95:87-90.
 11. Nawwar MAM, Barakat HH, Buddrust J, Linscheidt M. Alkaloidal, lignan and phenolic constituents of *Ephedra alata*. *Phytochemistry.* 1985; 24:878-879.
 12. O'Dowd NA, McCauley PG, Wilson G, Parnell JAN, Kavanaugh TAK, McConnell DJX. *Ephedra* Species: *In vitro* culture, micropropagation and the production of ephedrine and other alkaloids. *Medicinal and Aromatic plants.* 1998; (41):154-193.
 13. Parsaemehr A, Sargsyan E, Javidnia KA. Comparative Study of the Antibacterial, Antifungal and Antioxidant Activity and Total Content of Phenolic Compounds of Cell Cultures and Wild Plants of Three Endemic Species of *Ephedra*. *Molecules.* 2010; 15:1668-1678.
 14. Abourashed EA, El-Alfy AT, Khan IA, Walker L. *Ephedra* in perspective: A current review. *Phytother Res.* 2003; 17:703-712.
 15. Eberhardt MV, Lee CY, Liu RH. Antioxidant activity of fresh apples. *Nature.* 2000; 405:903-904.
 16. Amakura Y, Yoshimura M, Yamakami S, Yoshida T, Wakana D, Hyuga M *et al.* Characterization of Phenolic Constituents from *Ephedra* Herb Extract. *Molecules.* 2013; 18:5326-5334.
 17. Servil M, Montedoro G. Contribution of phenolic compounds in virgin olive oil quality. *European Journal of Lipid Science and Technology.* 2002; 104:602-613.
 18. Prakash A, Rigelhof F, Miller E. Antioxidant Activity, 2011. Available from World Wide Web: http://www.medlabs.com/downloads/antiox_acti_.pdf
 19. Kang TH, Pae HO, Yoo JC, Kim NY, Kim YC, Ko GI. Antiproliferative effect of alkaloids from *Sedum sarmentosum* on murine and human hepatoma cell line. *J Ethnopharmacol.* 2000; 70:177-182.
 20. Ruffa MJ, Ferraro G, Wagner ML, Calcagno ML, Campos RH, Cavallaro L. Cytotoxic effect of Argentine medicinal plant extracts on human hepatocellular carcinoma cell line. *J Ethnopharmacol.* 2002; 79(3):335-339.
 21. El-Baz FK, Hussein RA, Mahmoud K, Abdo SM. Cytotoxic activity of carotenoid rich fractions from *Haematococcus pluvialis* and *Dunaliella salina* microalgae and the identification of the phytoconstituents using LC-DAD/ESI-MS. *Phytotherapy Research.* 2018; 32:298-304.
 22. Mosmann T. Rapid colorimetric assays for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J Immunol Methods.* 1983; 65:55-63.
 23. Thabrew MI, Hughes RD, McFarlane IG. Screening of hepatoprotective plant components using a HepG2 cell cytotoxicity assay. *J Pharm Pharmacol.* 1997; 49:1132-1135.
 24. Ezzat MS, El-Hawary SS, El-Shabrawy AER, El-Shibani F. Evaluation of the phenolic and flavonoid contents, antimicrobial and cytotoxic activities of some plants growing in Al Jabal Al-Akhdar in Libya, *International Journal of Pharmacognosy and Phytochemical Research.* 2016; 8(7):1083-1087.
 25. Ghasemi M, Azarnia M, Jamali M, Mirabolghasemi G, Nazarian S, Naghizadeh MM *et al.* Protective effects of *Ephedra pachyclada* extract on mouse models of carbon tetrachloride-induced chronic and acute liver failure. *Tissue and Cell.* 2014; 46(1):78-85.
 26. Fan Y, Li J, Yin Q, Zhang Y, Xu H, Shi X *et al.* Effect of extractions from *Ephedra sinica* Stapf on hyperlipidemia in mice. *Experimental and therapeutic medicine.* 2015; 9(2): 619-625.
 27. Song M K, Um Jaeyoung, Jang H J & Lee B C. Beneficial effect of dietary *Ephedra sinica* on obesity and glucose intolerance in high-fat diet-fed mice. *Experimental and therapeutic medicine.* 2012; 3(4):707-712.
 28. Oshima N. Efficient preparation of Ephedrine alkaloids-free *Ephedra* herb extract and its antitumor effect and putative marker compound. [J] *Yakugaku Zasshi.* 2017; 137(2):173-177.
 29. Ramos S. Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. *J Nutr Biochem.* 2007; 18:427-442.
 30. Kanadaswami C, Lee L, Lee PH, Hwang J, Ke F, Huang Y *et al.* The antitumor activities of flavonoids. *In Vivo.* 2005; 19:895-909.
 31. Ren W, Qiao Z, Wang H, Zhu L, Zhang L. Flavonoids: promising anticancer agents. *Med Res Rev.* 2003; 23:519-534.